Diagnostic BioSystems

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Papanicolaou (PAP) Stain Kit

Catalog Number: KT038

**This data sheet is applicable to all sizes (volume) of product. Actual volume is indicated on vial.

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Intended Use
For In Vitro Diagnostic Use

Summary and Explanation
The Papanicolaou (PAP) Stain Kit is designed to differentiate between a variety of cells in vaginal smears for detection of vaginal, uterine and cervical cancer. In addition, this procedure is valuable for staining a variety of other bodily secretions and cell smears. The procedure was developed in the early 1940's by George Papanicolaou.

Nuclei: Blue
High Keratin Cells: Orange
Superficial Cells: Pink
Erythrocytes: Dark Pink
Parabasal Cells: Blue/Green
Intermediate Cells: Blue/Green
Metaplastic Cells: May contain both Blue/Green and Pink

Control Tissue
Tissues fixed in 10% formalin are suitable for use prior to paraffin embedding. Consult references (Kiernan, 1981: Sheehan & Hrapchak, 1980) for further details on specimen preparation.

1. Cut sections, usually 3 to 5 µm and pick the sections up on glass slides.
2. Bake the slides for at least 30 minutes at approximately 70°C.
3. Allow to cool.

Recommended Positive Control

1. Gynecological Smear
2. Any Superficial Cell Smear

Reagents Provided

Kit Contents	Volume	Storage
Modified Mayer's Hematoxylin	500 mL	15-30°C
OG-6 Solution	500 mL	15-30°C
EA-50 Stain Solution	500 mL	15-30°C

Storage and Handling
Do not use product after the expiration date printed on vial. If reagents are stored under conditions other than those specified here, they must be verified by the user. Diluted reagents should be used promptly.

Staining Procedure

1. Place slide in 95% alcohol for 5 minutes.
2. Place slide in 70% alcohol for 5 minutes.
3. Place slide in distilled water for 2 minutes.
4. Apply adequate Modified Mayer's Hematoxylin to completely cover cell smear and incubate for 5 minutes.
5. Rinse slide 1 time in distilled water to remove excess stain.
6. Rinse slide in tap water for 2 minutes.
7. Rinse slide in 2 changes of distilled water.
8. Dip slide several times in 95% alcohol and blot excess off.
9. Apply adequate OG-6 Stain Solution to completely cover cell smear to excess and incubate for 2 minutes.
10. Rinse slide gently using absolute alcohol.
11. Apply adequate EA-50 Stain Solution to completely cover cell smear to excess and incubate for 3 minutes.
12. Rinse slide gently using absolute alcohol.
13. Quickly dehydrate slide in 3 changes of absolute alcohol.
14. Clear slide and mount in synthetic resin.

Limitations of the Procedure

1. Histological staining is a multiple step diagnostic process that requires specialized training in the selection of the appropriate reagents, tissue selections, fixation, processing, preparation of the slide, and interpretation of the staining results.
2. Tissue staining is dependent on the handling and processing of the tissue prior to staining.
3. Improper fixation, freezing, thawing, washing, drying, heating, sectioning, or contamination with other tissues or fluids may produce artifacts or false negative results.
4. The clinical interpretation of any positive staining, or its absence, must be evaluated within the context of clinical history, morphology and other histopathological criteria. It is the responsibility of a qualified pathologist to be familiar with the special stain and methods used to produce the slide.
5. Staining must be performed in a certified licensed laboratory under the supervision of a pathologist who is responsible for reviewing the stained slides and assuring the adequacy of positive and negative controls.

Precautions

1. Consult local and/or state authorities with regard to recommended method of disposal.
2. Materials of human or animal origin should be handled as biohazardous materials and disposed of with proper precautions.
3. Avoid microbial contamination of reagents. Contamination could produce erroneous results.
4. This reagent may cause irritation. Avoid contact with eyes and mucous membranes.
5. If reagent contacts these areas, rinse with copious amounts of water.
6. Do not ingest or inhale any reagents.

Troubleshooting
If unexpected staining is observed which cannot be explained by variations in laboratory procedures and a problem is suspected, contact Diagnostic BioSystems Technical Support at (925) 484-3350, extension 2 or techsupport@dbiosys.com.

References

I. Papanicolaou, G.N. Atlas of Exfoliative Cytology, Harvard University Press, Cambridge, 1954.

Diagnostic BioSystems
6616 Owens Drive
Pleasanton, CA 94588
Tel: (925) 484 3350
www. dbiosys.com

CE

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EC REP

Emergo Europe
Prinsessegracht 20
2514 AP, The Hague
The Netherlands
Tel: (31) (0) 70 345 8570