

Instructions For Use KT033-IFI

Rev. Date: June 12, 2018

Revision: 6

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Copper Stain Kit (For Microwave)

Description: The Copper Stain Kit (For Microwave) is intended for the demonstration of

copper deposits in tissue sections.

Copper Deposits: Light Brown to Red

Nuclei: Blue

Uses/Limitations: For In-Vitro Diagnostic use only.

Histological applications.

Do not use past expiration date.

Use caution when handling these reagents.

Control Tissue: Fetal Liver or a known positive.

Availability/Contents:

Kit ContentsVolumeStorageRhodanine Solution (Stock)30 ml2-8°CAcetate Buffer Solution, pH 8.02 x 500 ml15-30°CHematoxylin, Mayer's (Lillie's Mod.) 125 ml15-30°C

Precautions: This product is a single-use, non-sterile, in vitro diagnostic device.

Keep away from open flame. Avoid contact with skin and eyes.

Harmful if swallowed.

Follow all Federal, State, and local regulations regarding disposal.

Use in chemical fume hood whenever possible.

Procedure (Standard):

Prepare Working Rhodanine Solution:

Combine:

4 ml Rhodanine Solution (Stock). Agitate Stock Solution immediately before adding to Acetate Buffer.

46 ml Acetate Buffer Solution, pH 8.0

- 1. Deparaffinize sections if necessary and hydrate to distilled water.
- 2. Place loosely capped staining jar containing Working Rhodanine in microwave and heat solution until warm but not hot.

Storage: 2° C 30° C

Mixed Storage Conditions.

Separate Contents.

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- 3. Place slide in warmed Working Rhodanine Solution and microwave at full power until solution is hot. Do not allow solution to boil.
- 4. Cap container, carefully agitate, and allow solution to cool on countertop to room temperature.
- 5. Examine slide microscopically and repeat heating/cooling cycle (steps 3 & 4) until desired staining intensity is achieved.
- 6. Rinse slide in 2 changes of Acetate Buffer Solution, pH 8.0 for 1 minute each.
- 7. Incubate slide in Hematoxylin, Mayer's (Lillie's Modification) for 5-10 seconds with agitation.
- 8. Rinse slide in 3 changes of Acetate Buffer Solution, pH 8.0 for 1 minute each.
- 9. Dehydrate slide in 3 changes of absolute alcohol.
- 10. Clear in 2 changes of xylene or xylene substitute, and mount in synthetic resin.

References:

- 1. Sheehan, DC., Hrapchak, BB. Theory and Practice of Histotechnology; 1980, page 230.
- 2. Lindquist, RR. Studies on the Pathogenesis of Hepatolenticular II: Cytochemical methods for the location of copper. Arch Pathol; 1969, Volume 87: page 370.

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