6616 Owens Drive, Pleasanton, CA 94588 U.S.A. - Tel. (925) 484-3350 - Fax (925) 484-3390 - www.dbiosys.com

Luxol Fast Blue Stain Kit

Description: The Luxol Fast Blue Stain Kit is designed for staining myelin/myelinated axons and Nissil substance on formalin fixed, paraffin-embedded tissue. This product is used for identifying the basic neuronal structure in brain or spinal cord sections.

Myelinated Fibers:	Blue
Nissil Substance:	Violet
Nerve Cells:	Violet

Uses/Limitations:
For In-Vitro Diagnostic use only.
Histological applications.
Do not use past expiration date.
Use caution when handling these reagents.

Control Tissue:
Cerebral Cortex
Spinal Cord

Availability/Contents:

Kit Contents	Volume	Storage
Cresyl Echt Violet Solution	125 ml	2-8°C
Luxol Fast Blue Solution	125 ml	15-30°C
Lithium Carbonate Solution (0.05%)	500 ml	15-30°C
Alcohol, Reagent (70%)	500 ml	15-30°C

Precautions: This product is a single-use, non-sterile, in vitro diagnostic device.
Avoid contact with skin and eyes.
May cause burns.
Harmful if swallowed.
Follow all Federal, State, and local regulations regarding disposal.
Use in chemical fume hood whenever possible.

Procedure (Standard):

1. Deparaffinize sections if necessary and hydrate to distilled water.
2. Incubate slide in Luxol Fast Blue Solution for 24 hours at room temperature or 2 hours at 60°C.
3. Rinse thoroughly in distilled water.
4. Differentiate section by dipping in Lithium Carbonate Solution (0.05%) several times (up to 20 seconds).
5. If needed, continue differentiation by repeatedly dipping in Alcohol, Reagent (70%) until gray-matter is colorless and white-matter remains blue.

Storage: 2° C 30° C

Mixed Storage Conditions.
Separate Contents.

Diagnostic BioSystems, Inc.
6616 Owens Drive
Pleasanton, CA 94588
U.S.A.

C V
P
Emergo Europe
Prinsessegracht 20
2514 AP The Hague, The Netherlands

6616 Owens Drive, Pleasanton, CA 94588 U.S.A. - Tel. (925) 484-3350 - Fax (925) 484-3390 - www.dbiosys.com

6. Rinse slide in distilled water.
7. Incubate slide in Cresyl Echt Violet (0.1%) for 2-5 minutes.
8. Rinse quickly in 1 change of distilled water.
9. Dehydrate quickly in 3 changes of absolute alcohol.
10. Clear as desired and mount in synthetic resin.

References:

1. Sheenan, D.C., Hrapchak, B.B. Theory and Practice of Histotechnology, 2nd Edition. Battelle Press, Columbus, OH. Page 262-264. 1980
2. Clark, G., et al., Staining Procedures. 4th Edition. Williams & Wilkins. Pages 146-147. 1981
3. Kluver, H., Barrera, E.A. A Method for the combined staining of cells and fibers in the nervous system. Journal of Neuropathology and Experimental Neurology, 1953, 12: pages 400-403.