

Trichrome Stain Kit (Modified Masson's)

Description: The Trichrome Stain Kit (Modified Masson's) is intended for use in the histological visualization of collagenous connective tissue fibers in formalin-fixed tissue sections.

Collagen:	Blue
Muscle Fibers:	Red
Nuclei:	Black/Blue


Uses/Limitations: For In-Vitro Diagnostic use only.
 Histological applications.
 Do not use past expiration date.
 Use caution when handling these reagents.
 Use of frozen sections or microwave during procedure may negatively affect staining.

Control Tissue: Lung
 Uterus
 Small Intestine
 Stomach

Availability/Contents:

<u>Kit Contents</u>	<u>Volume</u>	<u>Storage</u>
Bouin's Fluid	125 ml	15-30°C
Weigert's Iron, Hematoxylin (A)	125 ml	15-30°C
Weigert's Iron, Hematoxylin (B)	125 ml	15-30°C
Biebrich Scarlet / Acid Fuchsin Sol. Phosphomolybdic/Phosphotungstic	125ml	15-30°C
Acid Solution	125 ml	15-30°C
Aniline Blue Solution	125 ml	15-30°C
Acetic Acid Solution (1%)	125 ml	15-30°C

Precautions: This product is a single-use, non-sterile, in vitro diagnostic device.
 Keep away from open flame.
 Avoid contact with skin and eyes.
 Harmful if swallowed.
 Follow all Federal, State, and local regulations regarding disposal.
 Use in chemical fume hood whenever possible.

Storage: 15° C  30° C
**Store All Components At
Room Temperature.**



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Procedure (Standard):

1. Deparaffinize sections if necessary and hydrate to distilled water.
2. Preheat Bouin's Fluid in a water bath to 56° - 64° centigrade in a fume hood or very well ventilated area.
3. Place slides in preheated Bouin's Fluid for 60 minutes followed by a 10 minute cooling period.
4. Rinse slide in tap water until section is completely clear.
5. Rinse once in distilled water.
6. Mix equal parts of Weigert's (A) and Weigert's (B) and stain slide with working Weigert's Iron Hematoxylin for 5 minutes.
7. Rinse slide in running tap water for 2 minutes.
8. Apply Biebrich Scarlet / Acid Fuchsin Solution to slide for 15 minutes.
9. Rinse slide in distilled water.
10. Differentiate in Phosphomolybdic/Phosphotungstic Acid Solution for 10-15 minutes or until collagen is not red.
11. Without rinsing, apply Aniline Blue Solution to slide for 5-10 minutes.
12. Rinse slide in distilled water.
13. Apply Acetic Acid Solution (1%) to slide for 3-5 minutes.
14. Dehydrate very quickly in 2 changes of 95% Alcohol, followed by 2 changes of Absolute Alcohol.
15. Clear in Xylene or Xylene Substitute, and mount in synthetic resin.

References:

1. Sheehan, DC., Hrapchak, BB. Theory and Practice of Histotechnology; 1980, page 190.
2. A.F.I.P. Laboratory Methods in Histotechnology; 1992, pages 132-133.

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30° C

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