



# Instructions For Use

## KT 014-IFU

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### Fontana-Masson Stain Kit

(For Argentaftin Cells and Melanin)

#### Description and Principle

The Fontana-Masson Stain Kit is intended for use in the histological visualization of Melanin and other argentaftin substances. Substances that can bind silver and reduce it to a visible metallic form without a separate reducing agent are said to be "argentaftin". Fontana-Masson stain has been reported to be useful in the identification of Capsule-Deficient *Cryptococcus neoformans* and typical *Cryptococcus neoformans*. Argentaftin granules and Melanin are demonstrated by silver impregnation using an Ammoniacal Silver Solution.

#### Expected Results

Argentaftin Cell Granules:	Black
Melanin:	Black
Cryptococci Cell Wall:	Black
Nuclei:	Red
Cytoplasm:	Light Pink

#### Kit Contents

1. Gold Chloride Solution (0.2%)	2-8°C
2. Silver Nitrate Solution (10%)	2-8°C
3. Sodium Thiosulfate Solution (5%)	15-30°C
4. Nuclear Fast Red Solution	15-30°C

#### Storage

#### Suggested Controls (not provided)

Tissue containing Hair Follicles or Skin for Melanin. Small Intestine or Appendix for Argentaftin Granules.

#### Uses/Limitations

For In-Vitro Diagnostic use only.

Do not use if reagents become cloudy or precipitate

Do not use past expiration date.

Use caution when handling reagents.

Non-Sterile

Intended for FFPE sections cut at 5-10µm.

This procedure has not been optimized for frozen sections.

Frozen sections may require protocol modification.

#### Storage

Mixed storage conditions. Store according to individual label instructions.

#### Safety and Precautions

Please see current Safety Data Sheets (SDS) for this product and components GHS classification, pictograms, and full hazard/precautionary statements. If there is any serious incident that has occurred in relation to the device, please contact the manufacturer: Diagnostic BioSystems Technical Support at (925) 484-3350, extension 2 or techsupport@dbiosys.com. If required, please report to the Competent Authority of the Member State in which the user and/or patient is established.

#### Required but not Included

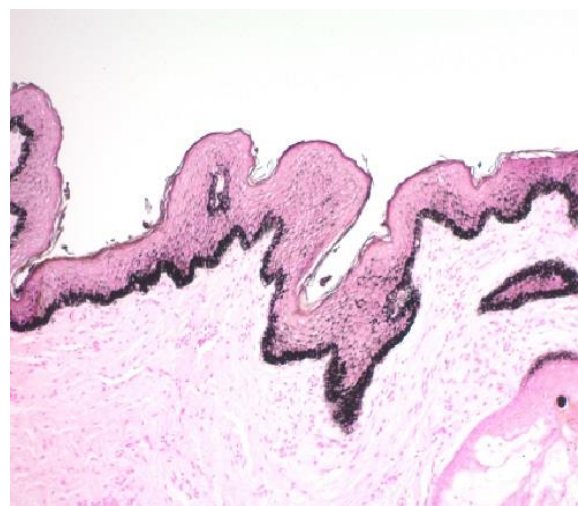
Concentrated Ammonium Hydroxide Solution (25-30%)

#### Important Notes

1. All glassware used in this procedure should be chemically cleaned and rinsed thoroughly in distilled water.

2. Do not use metal forceps to remove slides from reagents. Use plastic forceps only.

3. Equilibrate all reagents to room temperature prior to use.



Melanin in the basal layer of the epidermis of Human Skin stained with Fontana-Masson Stain. Magnification 100X

#### Preparation of Reagent Prior to Beginning:

Prepare Ammoniacal Silver Solution immediately prior to use.

In new or chemically cleaned glassware, mix 27ml Distilled/Deionized water with one vial of Silver Nitrate Solution (10%) and blend completely. Carefully add Concentrated Ammonium hydroxide (25-30%) (Not included) one drop at a time, swirling gently after each drop. Initially the mixture will turn dark brown and then gradually become transparent with a fine layer of sediment. The solution is ready for immediate use when all sediment dissolves.

#### Procedure

1. Deparaffinize sections if necessary and hydrate to distilled water.

2. Place freshly mixed Ammoniacal Silver Solution in a 58-60°C water bath and allow adequate time for temperature to equalize.

3. Incubate slide in warmed Ammoniacal Silver Solution for 30-60 minutes or until tissue section becomes yellowish/brown in color. (NOTE: Melanin typically stains in 30 minutes while Argentaftin granules stain in 50-60 minutes)

4. Rinse in 3 changes of distilled water.

5. Incubate slide in Gold Chloride Solution (0.2%) for 30 seconds.



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6. Rinse in 3 changes of distilled water.
7. Incubate slide in Sodium Thiosulfate Solution (5%) for 1-2 minutes.
8. Rinse for 2 minutes in running tap water followed by 2 changes of distilled water.
9. Incubate slide in Nuclear Fast Red Solution for 5 minutes.
10. Rinse for 2 minutes in running tap water followed by 2 changes of distilled water.
11. Dehydrate very quickly in 3 changes of absolute alcohol.
12. Clear, and mount in synthetic resin.

### References

1. Sheenan, D.C., Hrapchak, B.B. Theory and Practice of Histotechnology, 2nd Edition. Battelle Press, Columbus, OH.
2. Gaitanis, G., et al. Novel application of the Masson-Fontana Stain for Demonstrating Malassezia Species Melanin-Like Pigment Production In Vitro and in Clinical Specimens. Journal of Clinical Microbiology. 2005, August; 43(8): pages 4147-4151.
3. Kimura, M., et al. Fontana-Masson – stained tissue from culture-proven mycoses. Archives of Pathology & Laboratory Medicine. 1998, December; 122(12): page 11.
4. Lazcano, O., et al. Combined Fontana-Masson-Mucin staining of Cryptococcus neoformans. Archives of Pathology & Laboratory Medicine. 1991, November; 115(11): pages 1145-1149.
5. Ro, J.Y., et al. Advantage of Fontana-Masson stain in capsule-deficient cryptococcal infection. Archives of Pathology & Laboratory Medicine. 1987, January; 111(1): pages 53-57.



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