

# Instructions For Use KT 014-IFU

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1. All glassware used in this procedure should be chemically cleaned and rinsed thoroughly in distilled water.

2. Do not use metal forceps to remove slides from reagents. Use plastic forceps only.

3. Equilibrate all reagents to room temperature prior to use.



Melanin in the basal laver of the epidermis of Human Skin stained with Fontana-Masson Stain. Magnification 100X

# Preparation of Reagent Prior to Beginning:

Prepare Ammoniacal Silver Solution immediately prior to use.

In new or chemically cleaned glassware, mix 27ml Distilled/Deionized water with one vial of Silver Nitrate Solution (10%) and blend completely. Carefully add Concentrated Ammonium hydroxide (25-30%) (Not included) one drop at a time, swirling gently after each drop. Initially the mixture will turn dark brown and then gradually become transparent with a fine layer of sediment. The solution is ready for immediate use when all sediment dissolves.

## Procedure

1. Deparaffinize sections if necessary and hydrate to distilled water.

2. Place freshly mixed Ammoniacal Silver Solution in a 58-60°C water bath and allow adequate time for temperature to equalize.

3. Incubate slide in warmed Ammoniacal Silver Solution for 30-60 minutes or until tissue section becomes yellowish/brown in color. (NOTE: Melanin typically stains in 30 minutes while Argentaffin granules stain in 50-60 minutes)

4. Rinse in 3 changes of distilled water.

5. Incubate slide in Gold Chloride Solution (0.2%) for 30 seconds.



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Fontana-Masson Stain Kit

(For Argentaffin Cells and Melanin)

## **Description and Principle**

The Fontana-Masson Stain Kit is intended for use in the histological visualization of Melanin and other argentaffin substances. Substances that can bind silver and reduce it to a visible metallic form without a separate reducing agent are said to be "argentaffin". Fontana-Masson stain has been reported to be useful in the identification of Capsule-Deficient Cryptococcus neoformans and typical Cryptococcus neoformans. Argentaffin granules and Melanin are demonstrated by silver impregnation using an Ammoniacal Silver Solution.

## **Expected Results**

Argentaffin Cell Granules:	Black	
Melanin:	Black	
Cryptococci Cell Wall:	Black	
Nuclei:	Red	
Cytoplasm:	Light Pink	

Kit Contents	Volume	Storage
1. Gold Chloride Solution (0.2%)	125 ml	2-8°C
2. Silver Nitrate Solution (10%)	5 x 9ml	2-8°C
3. Sodium Thiosulfate Solution (5%)	125 ml	15-30°C
4. Nuclear Fast Red Solution	125 ml	15-30°C

## Suggested Controls (not provided)

Tissue containing Hair Follicles or Skin for Melanin. Small Intestine or Appendix for Argentaffin Granules.

# **Uses/Limitations**

For In-Vitro Diagnostic use only. Do not use if reagents become cloudy or precipitate Do not use past expiration date. Use caution when handling reagents. Non-Sterile Intended for FFPE sections cut at 5-10µm. This procedure has not been optimized for frozen sections. Frozen sections may require protocol modification.

## Storage

Mixed storage conditions. Store according to individual label instructions.

## Safety and Precautions

Please see current Safety Data Sheets (SDS) for this product and components GHS classification, pictograms, and full hazard/precautionary statements. If there is any serious incident that has occurred in relation to the device, please contact the manufacturer: Diagnostic BioSystems Technical Support at (925) 484-3350, extension 2 or

techsupport@dbiosys.com. If required, please report to the Competent Authority of the Member State in which the user and/or patient is established.

## **Required but not Included**

Concentrated Ammonium Hydroxide Solution (25-30%)

# **Important Notes**



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6. Rinse in 3 changes of distilled water.

7. Incubate slide in Sodium Thiosulfate Solution (5%) for 1-2 minutes.

8. Rinse for 2 minutes in running tap water followed by 2 changes of distilled water.

9. Incubate slide in Nuclear Fast Red Solution for 5 minutes.

10. Rinse for 2 minutes in running tap water followed by 2 changes of distilled water.

11. Dehydrate very quickly in 3 changes of absolute alcohol.

12. Clear, and mount in synthetic resin.

#### References

1. Sheenan, D.C., Hrapchak, B.B. Theory and Practice of Histotechnology, 2nd Edition. Battelle Press, Columbus, OH.

2. Gaitanis, G., et al. Novel application of the Masson-Fontana Stain for Demonstrating Malassezia Species Melanin-Like Pigment Production In Vitro and in Clinical Specimens. Journal of Clinical Microbiology. 2005, August; 43(8): pages 4147-4151.

3. Kimura, M., et al. Fontana-Masson – stained tissue from culture-proven mycoses. Archives of Pathology & Laboratory Medicine. 1998, December; 122(12): page 11. 4.Lazcano, O., et al. Combined Fontana-Masson-Mucin staining of Cryptococcus

neoformans. Archives of Pathology & Laboratory Medicine. 1991, November; 115(11): pages 1145-1149.

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